

Asymmetric Reduction of Karahanaenone with Various Microorganisms

Mitsuo Miyazawa,* Keisuke Tsuruno and Hiromu Kameoka

Department of Applied Chemistry, Faculty of Science and Engineering, Kinki University, Kowakae,
 Higashiosaka-shi, Osaka 577, Japan.

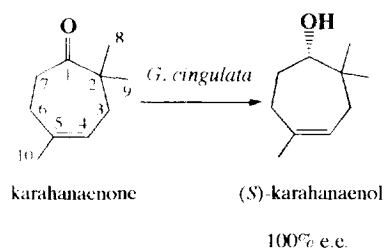
Abstract—Both enantiomers of karahanaenol have been prepared by microbial asymmetric reduction of karahanaenone. The (*S*)-enantiomer with high enantiomeric excess was obtained by *Glomerella cingulata*, *Aspergillus niger*, *Streptomyces aureofaciens* and *Bacillus subtilis* respectively. In contrast, the (*R*)-enantiomer of karahanaenol was obtained with *Fusarium solani*.

Karahanaenone is a monoterpene present in hop oil¹. This unique and peculiar compound with its fragrance and flavour is one of the most important compounds for the flavouring industry². Our main objective is to make a compound with a unique fragrance and flavour and which possesses a fruity and a woody odor. The reduction of karahanaenone by NaBH₄ has been reported³. Chemical and biological methods have been applied to the asymmetric reduction of karahanaenone into karahanaenol however the enantioselectivity was poor. In this paper, we report a novel method for reducing karahanaenone with various microorganisms into karahanaenol with excellent enantioselectivity.

We selected the following fungi *Aspergillus niger* IFO 4414, *Glomerella cingulata*⁴, *Fusarium solani*⁴ and *Rhizoctonia solani*⁴, and the bacterium *Bacillus subtilis* IFO 3936, and actinomyces *Streptomyces aureofaciens* IFO 3187. All the microorganisms reduced karahanaenone during a period of 24hrs. The results are summarized as following in the table below.

Table : Microbiological Reduction of karahanaenone

	$[\alpha]_D^{20}$	ee	Conf.	Yield
<i>Glomerella cingulata</i>	+33	100%	S	98%
<i>Aspergillus niger</i>	+33	100%	S	98%
<i>Rhizoctonia solani</i>	+28	70%	S	97%
<i>Streptomyces aureofaciens</i>	+33	100%	S	96%
<i>Bacillus subtilis</i>	+33	100%	S	98%
<i>Fusarium solani</i>	-28	70%	R	80%



The absolute configuration of the secondary alcohol was determined from esters obtained by the reaction of each karahanaenol (*S*)-(-) and (*R*)-(+)-(α)-MTPA chlorides according to Mosher's methods^{5,6}. Enantiomerically pure karahanaenol has not been described in the literature before.

Both enantiomers of karahanaenol have a mint like odor. The (*S*)-enantiomer had a notably fresher odor than the (*R*).

Experimental

Reduction on analytical scale. Fungi case: Mycelia were respectively transplanted to the culture medium (15ml in a 50ml Petri dish) and furthermore incubated at 27°C under a static situation for 3days. After the growth of the fungus, the substrate was added directly to each medium (7.5mg/15ml) and further incubated at the same condition for 24h. Bacteria and Actinomyces case: Mycelia were then transplanted to the culture medium (100ml in a 200ml Erlenmeyer flask) and stirred for 3days at 28°C. After the growth of the fungus, the substrate was added directly to each medium (50mg/100ml) and were further incubated at the same condition for 24h. Nutrient broth for microbial transformation: *A. niger*, *F. solani*, *G. cingulata*: polypeptone 5g, KCl 0.5g, MgSO₄·7H₂O 0.01g, succharose 15g, glucose 15g, H₂O 1l. *R. solani*: glucose 50g, KNO₃ 10g, K₂HPO₄ 5g, MgSO₄·7H₂O 2.5g, FeCl₃·6H₂O 0.02g, H₂O 1l. *B. subtilis*: polypeptone 5g, Yeast extract 0.2g, MgSO₄·7H₂O 0.1g, H₂O 1l. *S. aureofaciens*: oatmeal 5g, H₂O 1l. Aliquots were withdrawing periodically and monitored by GLC on a chiral column : CHIROMPACK WCOT Fused silica Cp-Cyclodextrin-B-236-M-19(50m×0.25mm , N₂ flow rate 1ml/min).

Reduction on preparative scale. Karahanenone was added to the resulting mycelia of the grown cell by a process similar to the analytical scale. After 24hrs, the reaction mixture was extracted with dichloromethane with a continuous liquid-liquid extractor and concentrated under reduced pressure. The products were purified by column chromatography (silica gel, hexane/ethyl acetate ratio) to give the enantiomerically pure alcohol. Optical rotations were measured on a Japan Spectroscopic CO. LTD DIP-140 in CHCl₃. The enantiomeric excesses are determined by GLC on chiral column [retention time (min) of alcohols : (*S*)-13.7, (*R*)-13.4].

Determination of absolute configuration of (-) karahanaenol. [$\Delta \delta$ = δ (-)-MTPA - δ (+)-MTPA : (C1) proton; 4.88-4.91=-0.03, (C2) gem Methyl; 0.93-0.86=+0.07, 0.83-0.81=+0.02 (C4) proton; 5.35-5.34=+0.01, (C5) Methyl; 1.71-1.74=-0.03, (C6) proton; 2.08-2.13=-0.05, 1.95-2.01=-0.06].

References and Notes

1. Naya, Y. and Kotake, M. *Tetrahedron Lett.* **1968**, 13, 16-45.
2. Nomura, M., Inoue, T. and Fujihara, Y. *Nippon Nogeikagaku Kaishi.* **1992**, 66(6), 999.
3. Miyawaki, H., Yugawa, C. and Yumoto, I. Jpn. Patent 61,218,543, **1986**.
4. The strain stored in Gifu University (Japan) was used in this study.
5. Dale, J. A., Dull, D. L. and Mosher, H. S. *J. Org. Chem.* **1969**, 34(9), 2543.
6. Ohtani, I., Kusumi, T., Kashman, Y. and Kakisawa, H. *J. Am. Chem. Soc.* **1991**, 113, 4092.

(Received in Japan 14 July 1995; accepted 23 August 1995)